

Revision Date: Sept-17-2023

IF Imaging of Cell Surface Epitopes

Cell Plating

Begin with a cell suspension of 0.5e6 cells/mL - 1.0e6 cells/mL

Antibody Staining

- 1. Pellet and wash cells 3x with PBS +/+
- 2. Dilute Cell Surface Bio Primary Antibody in blocking buffer (10% NGS recommended) to desired concentration (1ug/mL recommended)
- 3. Use 1-4% Paraformaldehyde (PFA) to fix the cells
- 4. Pellet and wash cells 3x with PBS +/+
- 5. Incubate cells with Primary Antibody for 1 hour
- 6. Pellet and wash Cells 2x with PBS +/+
- 7. Dilute secondary antibody conjugated with fluorophore of your choice in blocking buffer.
- 8. Incubate cells with diluted secondary for 30 minutes
- 9. Pellet and wash cells 3x with PBS +/+
- 10. Use 1-4% paraformaldehyde (PFA) to fix the cells
- 11. Pellet and wash cells 3x with PBS+/+
- 12. Add DAPI and incubate for 5 minutes
- 13. Pellet and wash with PBS +/+ 2X
- 14. Add PBS +/+ and Image using fluorescent imager

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Material

- Cell Surface Bio primary antibody
- Cells
- Blocking buffer (NGS)
- PBS +/+
- Fluorophore conjugated anti-mouse secondary antibody
- Paraformaldehyde (PFA)
- DAPI
- Fluorescent imager